

Formulation of Polyherbal cream and evaluation of their antimicrobial potential Anushree Jyotishi¹, Pradeep Kumar Mohanty, Vishal Shrivastava, Akhlesh Kumar Singhai and Sourabh Malviya²

1, School of Pharmacy, LNCT University, Kolar Road, Bhopal, (M.P)

2, Minor Forest Produce Processing and Research Centre, Barkheda Pathani, Bhopal, (M.P)

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Abstract

In recent years, there has been a progressive gradual development in the use of medicinal plants in developing nations, owing to the fact those herbal medicines are safe and have few side effects, especially when compared to synthetic pharmaceuticals. As the occurrence of unwanted side effects appears to be less common with herbal medications compared to allopathic treatments, the usage of herbal drugs is increasing, resulting in a rapid increase in the number of herbal drug makers. Since they are typically less expensive than synthetic medications People have been increasingly using herbal medicines during the last few decades. The present study was commenced to evaluate the quantitative profiling and antimicrobial activity of cream prepared from extracts of traditional plants like Kanthkari, Rasana, and Nirgundi. In the present study, a cream was formulated which comprised of an extract of plant materials. The dried plant materials of Kanthkari, Rasana, and Nirgundi were individually extracted using the maceration method.

These three different types of extracts were used for the preparation of cream and evaluated for the HPTLC profiling and antimicrobial activity. The HPTLC profiling of the extract and formulation was performed on pre-coated silica gel Aluminum Plate 60F-254 using Toluene: Ethylacetate: Formic Acid (8:1.5:0.5v/v/v) as mobile phase and a further quantity of Quercetin was evaluated in extract and formulation. The antimicrobial activity of prepared cream was evaluated against *Escherichia coli* and *Staphylococcus aureus* by disk diffusion method using reference disk of antibiotics. The result of antimicrobial the activity was determined by comparing the results of the zone of inhibition of formulation and solvent extract. The formulation showed significant activity against the tested bacterial pathogens. The disk diffusion method was used to assess the antibacterial activity of methanolic extract using a reference disk of antibiotics. The R_f value and spectral scanning of standard Quercetin (R_f 0.22) match with the values of extract and formulation indicate the presence of Quercetin in extract and formulation. Considering the ability of the golden treasure present in Kanthkari, Rasna, and Nirgundi. The Quality Control profiling of formulation and extract was performed by HPTLC, and antimicrobial activity was established by disk diffusion method. Formulated cream can use as a potential antibacterial against various bacterial infections.

Key-words: Polyherbal cream, Antimicrobial activity, Formulation

Introduction

Topical skin infections commonly occur and often present therapeutic challenges to practitioners, despite the numerous existing antibacterial agents available today. The necessity for developing new antibacterial means has increased significantly due to growing concerns regarding multidrug-resistant bacterial, viral, and fungal strains[1-4]. Common examples for topical skin infections include diaper rash, cold sores, and tinea (also called pityriasis) versicolor. Topical delivery is

the application of a drugcontaining formulation to the skin to treat cutaneous disorders (e.g. acne) or cutaneous symptoms of a general disease (e.g. psoriasis) with the goal of limiting the drug's pharmacological or other action to the skin's surface or within the skin.

*Corresponding Author

E.mail: anushree121jyo@gmail.com

Its development is multifactorial, including skin wetness, friction, skin irritants, and pH change, which favors the growth of microorganisms including *Candida*, *Staphylococcus*, and *Streptococcus*[5].

India has a long history of medicinal herbs, as well as a variety of ancient medical systems. Traditional medicine, which is widely practiced in India, includes a vast variety of plants with varied therapeutic and pharmacological significance, and so provides a priceless reservoir of new bioactive compounds. In recent years, there has been a greater emphasis on plant research around the world, and data has accumulated to illustrate the enormous potential of medicinal plants employed in diverse traditional systems. From a drug development standpoint, plant chemical compounds will hit the drug target at certain places and rule over the synthetic substance[6]. As a result, plant chemical components are one of the greatest sources for most significant new medication discoveries. Traditional medicines are still used by more than 60% of the world's population to cure various diseases rather than allopathic medicine systems[7]. Traditional medicinal plants including Kanthkari, Rasana, and Nirgundi are well-known for their therapeutic properties. Different parts of them have been used by Indian traditional healers to treat a range of diseases, such as Rasana, the plant is used for the inflammation and bronchitis, psoriasis, cough and piles. It is also used as antipyretic, analgesic, dyspepsia, rheumatoid arthritis, bitter, laxative

and nervetonic. The decoction of the plant is used to prevent the swelling of joints in arthritis, rheumatism and neurological disorders [8,9] and Nirgundi possesses analgesic, anti-parasitic, antibacterial, antipyretic, antihistaminic, analgesic, insecticidal, CNS depressant activity, and anti-fertility[10]. Kanthkari, It is commonly used in bronchial asthma, cough, worms, etc several other medicinal uses like anthelmintic, antipyretic, anti-inflammatory, antitumor, cytotoxic activities, anti-asthmatic, antispasmodic, and hypotensive[11,12]. The aim of the present study was to formulate topical creams containing extract of Rasna, Kanthkari and Nirgundi and to evaluate the *in vitro* antibacterial activity in the formulations against *Escherichia coli* and *Staphylococcus aureus*.

Experimental

Collection of plants sample



The fresh leaves and roots of Kanthkari, Rasana, and Nirgundi were collected from Minor Forest Produce Processing and Research Centre, Bhopal. The plant was identified and authenticated by Ms. Mahima of Vedanta Testing and Research laboratory, Bhopal.

Pharmacognostic study: The sample was evaluated on different parameters like Macroscopic, Microscopic, Physiochemical and Phytochemical to ensure the quality, authenticity of the plant material.

Table 1: Pharmacognostic investigation

| Pharmacognostic investigation of Rasna | |
|--|-----------------------------|
| BotanicalName | <i>Pluchea lanceolata</i> |
| Family | Asteraceae |
| CommonName: | Rasna |
| Macroscopic Examination | |
| Condition | Dried |
| Colour | Darkgreen |
| Odour | Characteristic |
| Taste | SlightlyastringentandBitter |

| | |
|--|--|
| Leaves | Leaves simple 2-5 cm long, 0.5 to 2cm broad, brittle, sessile obtuselanceolate, reticulate, midrib more distinct at the lower side, lateral veins somewhat running straight parallel to the midrib; entire toothed near the apex, acute base symmetrical both surface pubescent. |
| Microscopic Examination | <div data-bbox="889 478 1300 953" data-label="Image"> </div> <p data-bbox="992 957 1133 989">Figure 1.1:</p> <p data-bbox="597 993 1528 1157">TS passing through the mid rib shows one or occasionally two centrally located meristless encircled by thick walled, lignified, dorsiventrally well-developed sclerenchyma sheath, radially arranged xylem vessel and well developed phloem tissue; collenchyma tissue, almost reaching up to the meristele lies underneath both the epidermis.</p> |
| Pharmacognostic investigation of Nirgundi | |
| BotanicalName | <i>Vitexnegundo</i> |
| Family | Verbenaceae |
| CommonName | Nirgundi leaves |
| Macroscopic Examination | |
| Condition | Dried |
| Colour | Darkgreen |
| Odour | Aromatic |
| Taste | Bitter |
| Leaves | Fresh leaf, leathery, palmately trifoliate to Penta foliate, the middle leaflet largest in size 6 to 12cm long 1.5 to 2cm wide with 1 to 2cm long petiole at base. Rachis long, leaflets, lanceolate, acute, entire or rarely crenate, pubescent and dark green on upper surface, tomentose and whitish on lower surface. |

| | |
|---|--|
| <p>Microscopic Examination</p> | <p>Figure 1.2:</p>  <p>TS of leaflet passing through the midrib is strongly convex on its lower side and somewhat flattened on its upper side, with an arc of meristele in the centre and narrow is bilaterallamina on its either side; the entire lower surface of the leaf pubescent. Mesophyll region of the lamina shows 2-5 compact rows of the palisade cells discontinuous over the midrib region but getting slightly narrow.</p> |
| <p align="center">Pharmacognostic investigation of Kantakari</p> | |
| <p>Botanical Name</p> | <p><i>Solanum virginianum</i> Linn.</p> |
| <p>Family</p> | <p>Solanaceae</p> |
| <p>Macroscopic Examination</p> | |
| <p>Condition</p> | <p>Dried</p> |
| <p>Colour</p> | <p>Yellowish green</p> |
| <p>Odour</p> | <p>Characteristic</p> |
| <p>Taste</p> | <p>Characteristic</p> |
| <p>Leaves</p> | <p>Brittle ovate oblong or rarely elliptic in shape 2 -12 cm in length, 2.5-5 cm in width, sinuate or sub pinnatifid, sub acute, pubescent; midrib and lateral veins shows prominent sharp prickles; petiole cylindrical, 1 -2 cm in length, pubescent with spines and longitudinally running furrowed upper side.</p> |
| <p>Microscopic Examination</p> |  <p>Figure 1.3:</p> <p>TS of leaf passing through midrib is dorsiventrally convex shows a narrow collenchymatous band underneath both the epidermis, a centrally located arc of biocollateral vascular bundle and two lateral narrow dorsiventral laminar extensions</p> |

Physicochemical analysis: Physicochemical analysis such as Percentage of ash values and extractive values were determined according to the Ayurvedic Pharmacopoeia of India [13].

Table 2: Physicochemical analysis

| Experimental Results | | | | |
|----------------------|------------------------------------|-------------|----------|-----------|
| S.No | Physicochemical parameters (% w/w) | Rasna Patti | Nirgundi | Kanthkari |
| 1 | Total ash | 12% | 2.6 % | 6.2% |
| 2 | Acid-insoluble ash | 1.5% | 0.62% | 1.6% |
| 3 | Alcohol-soluble extractive | 28.6 % | 12.1% | 12.2 % |
| 4 | Water-soluble extractive | 26.2% | 8.2% | 11% |

Preparation of Extract:

The two fraction of plant sample was prepared using methanol and Triple distilled water.

Methanolic Extract- Coarsely powdered 100g of drug was macerated and extracted with 250 ml methanol at room temperature for 7 days and the extract was concentrated, frozen and lyophilized by lyophilizer.

Aqueous Extract- Coarsely powdered 100g of powdered drug was macerated and extracted with at room temperature for 7 days and the extract was concentrated by placed in oven at not more than 40°C for about 24 hours [14].

Preliminary Phytochemical Investigation.

The plant extracts were assessed for the existence of the phytochemical classes as described by Trease and Evans (2002), and Harborne (1973). Investigations on the preliminary phytochemical screening of Kanthkari, Rasana, and Nirgundi extracts revealed the presence of phenols, flavonoids, tannins, saponins, alkaloids and carbohydrates in methanolic and aqueous extracts respectively [15,16].

Table 3: Preliminary Phytochemical Investigation.

| Chemical Constituents | Methanolic extract | | | Water Extract | | |
|--------------------------|--------------------|----------|-----------|---------------|----------|-----------|
| | Rasna Patti | Nirgundi | Kanthkari | Rasna Patti | Nirgundi | Kanthkari |
| Phenols | - | + | + | - | + | + |
| Glycosides | + | + | + | + | + | + |
| Flavonoids | + | - | + | + | - | + |
| Tannins | + | + | + | + | + | + |
| Saponins | + | + | + | + | + | + |
| Alkaloids | - | + | + | - | + | + |
| Carbohydrates | + | - | + | + | - | + |
| Proteins and amino acids | + | - | + | + | - | + |

Formulation

The ingredients were weighed as per the details given in Table below. The oily phase (Part A) that

consisted of the emulsifier (stearic acid) and other oil soluble components was heated to 75±1° C on a water bath shaker (with constant stirring. The water soluble components were added to water

(Part B) and heated to the same temperature followed by addition of Rasna extract, Nirgundi extract, Kanthkari extract, methyl and propyl paraben with continuous stirring. To the heated aqueous mixture, oily phase was incorporated

with continuous stirring on magnetic stirrer (Jindal Scientific Industries Pvt. Ltd) until the emulsion cooled down[17].

Table 4: Formulation of Herbal Cream

| PART A (Oily Phase) | | PART B (Aqueous Phase) | |
|-----------------------|-------|------------------------|----------|
| Ingredient | %w/w | Ingredient | %w/w |
| Light liquid paraffin | 27.85 | Triethanolamine | 1.71 |
| Stearic acid | 8.57 | Glycerin | 10.71 |
| Glyceryl monostearate | 7.50 | Methyl paraben | 0.21 |
| Cetostearyl alcohol | 4.28 | Propyl paraben | 0.21 |
| Microcrystalline wax | 0.21 | Rasna extract | 2.10 |
| Hard paraffin wax | 1.00 | Nirgundi extract | 2.10 |
| | | Kanthkari Extract | 2.10 |
| | | Distilled water | q.s.100% |

Evaluation of cream

The formulation properties of the cream were studied visually and characteristics like physical

state, colour, odour and overall appearance was recorded[18].

Table 5: Evaluation Parameter of cream

| Parameter | Result |
|---------------|--------------------------------|
| Color | Dark Green |
| Odour | Sweet |
| Consistency | Smooth |
| State | Stable Semi Solid |
| pH | 6.52±0.20 |
| Spreadability | 28.89 ± 1.11g.cm/sec |
| Viscosity | 64000 cps (at least rpm of 10) |

Estimation of Quercetin by HPTLC

Standard stocksolution:

Standard Quercetin was procured from Natural Remedies, Bangalore, India. 10mg/10ml Quercetinas standard solution was prepared in methanol.

Mobile phase: Mobile phase consists of a mixture of Toluene: Ethylacetate: Formic Acid (8:1.5:0.5v/v/v).

Preparation of test solution for Kanthkari:

10 grams of coarsely powdered drug were taken in 250 ml stoppered conical flask and extract with 100 ml alcohol for 24 hours by maceration with occasional shaking. The extract was decanted and

made up to 100 ml in volumetric flask. The solution is filtered through Whatman filter paper no.41, 25 ml of the extract was taken from stock solution and dried on a water bath. Extract the dried bark with n-hexane (2x 5ml). Concentrate the pooled n- hexane extract to 5ml.

Preparation of test solution for Rasna:

10 grams of coarsely powdered drug were taken in 250 ml stoppered conical flask and extract with 100 ml alcohol for 24 hours by maceration with occasional shaking. The extract was decanted and made up to 100 ml in volumetric flask. The solution is filtered through Whatman filter paper no.41, 25 ml of the extract was taken from stock solution and dried on a water bath. The dried mark

was extracted with Petroleum ether 60-80°C (2x5ml). Concentrate the pooled Petroleum ether 60-80 °C extract to 5ml.

Preparation of test solution for Nirgundi:

Take 10 grams of coarsely powdered drug were taken in 250 ml stoppered conical flask and extract with 100 ml alcohol for 24 hours by maceration with occasional shaking. The extract was decanted and made up to 100 ml in volumetric flask. The solution was filtered through Whatman filter paper no.41, 25 ml of the extract was taken from stock solution and concentrate on water bath to 5ml.

Instrumentation and chromatographic conditions

Total 12 bands were spotted with width of 8mm by a Camag 100 microliter syringe on pre-coated silica gel aluminum Plate 60F-252 (20 cm × 10 cm with 0.2 mm thickness, E. Merck, Germany) using a Camag Linomat 5 (Switzerland). A constant application rate of 150nl/s was employed and space between two bands was 8 mm. The slit dimension was kept 6 mm × 0.20 mm

micro; 100nm/s scanning speed was employed. The mobile phase consisted of Toluene: Ethylacetate: Formic Acid (8:1.5:0.5v/v/v). Linear ascending development was carried out in a twin trough glass chamber saturated with the mobile phase. The optimized chamber saturation time for the mobile phase was 20 min at room temperature. The length of the chromatogram run was approximately 70 mm. Subsequent to the development; the TLC plate was dried in a current of air with the help of an air dryer. Densitometric scanning was performed on Camag TLC scanner in the absorbance mode at 252 nm. The source of radiation utilized was deuterium lamp emitting a continuous UV spectrum between 200 and 400 nm. The content of Quercetin was estimated in the extract and dosage forms by spotting 10µl solution of Kanthkari and 2 µl solution of the Rasna, Nirgundi, and formulation sample on a TLC plate. The TLC plate was developed to a distance of 6 cm from the point of application. TLC plate was air dried and scanned on a Camag TLC Scanner in its absorbance mode at 252 nm.

Figure 2: Scanning of TLC plate via Camag TLC Visualizer

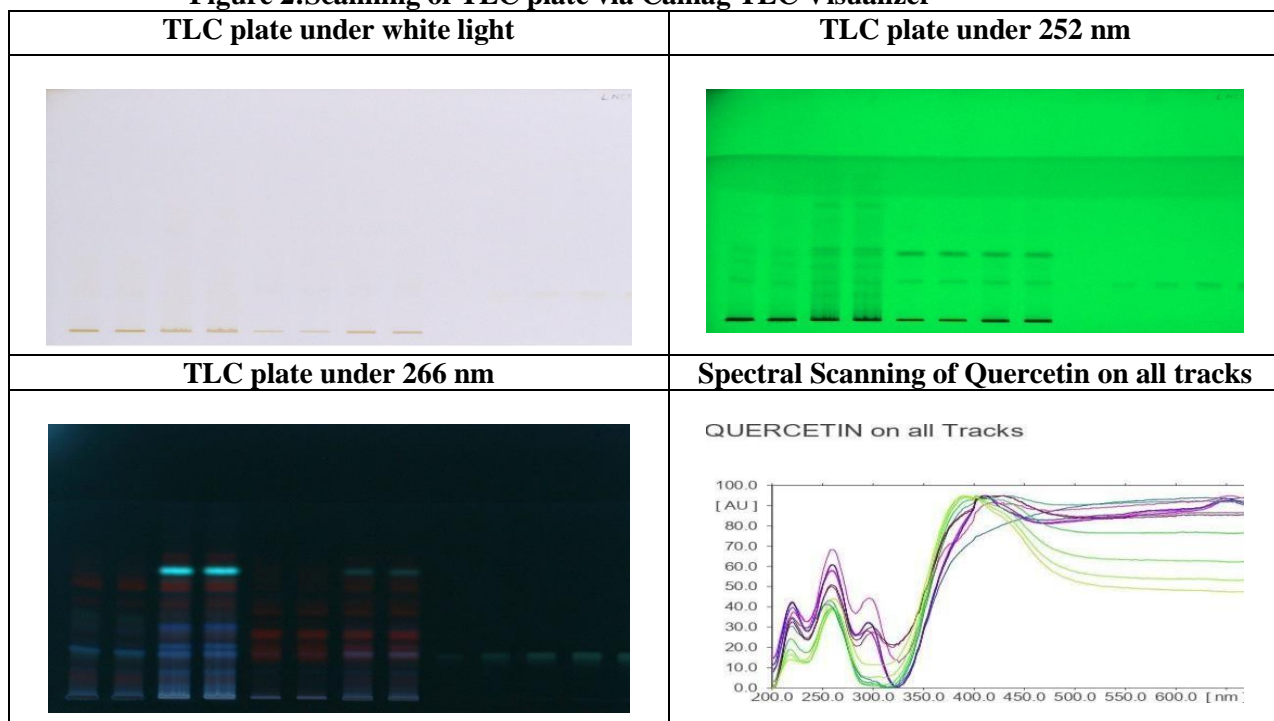


Table 6: Rf value of extract and formulation indicate the presence of Quercetin

| Tracks Number | Name | Rf value | Peak area |
|---------------|--------------------|----------|-----------|
| 1. | Rasnapatti extract | 0.25 | 1012.1 |
| 2. | Rasnapatti extract | 0.22 | 1122.9 |
| 3. | Kantkari extract | 0.22 | 681.9 |
| 4. | Kantkari extract | 0.22 | 685.6 |
| 5. | Nirgundi extract | 0.22 | 1521.9 |
| 6. | Nirgundi extract | 0.22 | 1226.2 |
| 7. | Formulation | 0.22 | 1266.2 |
| 8. | Formulation | 0.22 | 1209.2 |
| 9. | Standard 1 | 0.22 | 228.2 |
| 10. | Standard 2 | 0.22 | 822.2 |
| 11. | Standard 2 | 0.22 | 1512.6 |
| 12. | Standard 2 | 0.22 | 2100.0 |
| 13. | Standard 5 | 0.22 | 2622.8 |
| 14. | Standard 6 | 0.22 | 2265.6 |

Estimation of Quercetin

The content of Quercetin was estimated in the extract and dosage forms by spotting 10µl solution of Kanthkari and 2µl solution of the Rasna, Nirgundi and formulations sample on a TLC plate. The TLC plate was developed to a distance of 6 cm from the point of application. TLC plate was air dried and scanned on a Camag TLC Scanner in its absorbance mode at 252 nm. The content of Quercetin was calculated by linear regression, and mean percentages were calculated from six replicate experiments.

| Substance: QUERCETIN @ 254 nm | | | | | | | | | |
|-------------------------------|------|--------|------------|------------------------|---------|-------------|---------|-----------------|--|
| Regression via area: | | Linear | | Y = -392.3 + 3.177 * X | | r = 0.99983 | | sdv = 1.35 | |
| Track | Vial | Rf | Amount | Height | X(Calc) | Area | X(Calc) | SampleID/Remark | |
| 1 | 1 | 0.25 | | | | 1013.10 | 442.38 | RASNAPATTI | |
| 2 | 1 | 0.23 | | | | 1039.00 | 450.54 | RASNAPATTI | |
| 3 | 2 | 0.23 | | | | 770.36 | 365.98 | KANHTKARI | |
| 4 | 2 | 0.24 | | | | 685.70 | 339.33 | KANHTKARI | |
| 5 | 3 | 0.24 | | | | 1521.91 | 652.54 | NIRGUNDI | |
| 6 | 3 | 0.24 | | | | 1402.79 | 565.05 | NIRGUNDI | |
| 7 | 4 | 0.24 | | | | 1266.36 | 522.10 | F1 | |
| 8 | 4 | 0.24 | | | | 1264.05 | 521.38 | F1 | |
| 9 | 5 | 0.23 | 200.00 ng | | | 245.32 | | | |
| 10 | 5 | 0.23 | 400.00 ng | | | 488.66 | | | |
| 11 | 5 | 0.23 | 600.00 ng | | | 732.00 | | | |
| 12 | 5 | 0.23 | 800.00 ng | | | 975.34 | | | |
| 13 | 5 | 0.23 | 1000.00 ng | | | 1218.68 | | | |
| 14 | 5 | 0.23 | 1.200 µg | | | 2765.36 | | | |
| | | | | | | 3439.07 | | | |

| Calibration results per Analysis | | | | | | | | | |
|----------------------------------|------|-------------|--------|---|------------|--|--|--|--|
| Sample from vial 1: RASNAPATTI | | | | | | | | | |
| Result via area | | | | | | | | | |
| Substance | Rf | X(average) | CV [%] | n | Regression | | | | |
| QUERCETIN | 0.24 | 446.46 ng | 1.291 | 2 | Linear | | | | |
| Sample from vial 2: KANHTKARI | | | | | | | | | |
| Result via area | | | | | | | | | |
| Substance | Rf | X(average) | CV [%] | n | Regression | | | | |
| QUERCETIN | 0.23 | 352.65 ng | 5.344 | 2 | Linear | | | | |
| AutoGenerated3 | 0.27 | 0.0 unknown | 0.000 | 0 | Linear | | | | |
| Sample from vial 3: NIRGUNDI | | | | | | | | | |
| Result via area | | | | | | | | | |
| Substance | Rf | X(average) | CV [%] | n | Regression | | | | |
| QUERCETIN | 0.24 | 583.80 ng | 4.541 | 2 | Linear | | | | |
| Sample from vial 4: F1 | | | | | | | | | |
| Result via area | | | | | | | | | |
| Substance | Rf | X(average) | CV [%] | n | Regression | | | | |
| QUERCETIN | 0.24 | 521.74 ng | 0.099 | 2 | Linear | | | | |

Antibacterial activity of plant extracts and formulation:

Anti-bacterial analysis

The anti-bacterial analysis was carried out using the methanolic extract of plants to assess the antibacterial activity. Antibacterial activity was analyzed against *Escherichia Coli* and *Staphylococcus aureus*. The extract was prepared as in the standard procedure. 1mg of dried extract was weighed and suspended in 1ml of dimethylsulphoxide (DMSO) to make a stock solution of 1mg/ml dose. Antibacterial activity was determined by the disc diffusion method to test the inhibitory zone formed by leaf extract. The sterile discs were taken and impregnated with the different concentrations of leaf extract then placed on nutrient agar media plates containing bacterial cultures and incubated at 26°C for 22 hrs. After incubation, the antibacterial activity of methanolic leaf extracts against the bacterial strains was

assessed by measuring the diameter of the inhibition zone formed [19].

Disk Diffusion Method

Different concentrations of herbal drug are compared with standard antibiotics, as recommended in the CLSI (Clinical and Laboratory Standards Institute) guidelines. The disk of antibiotics contains:

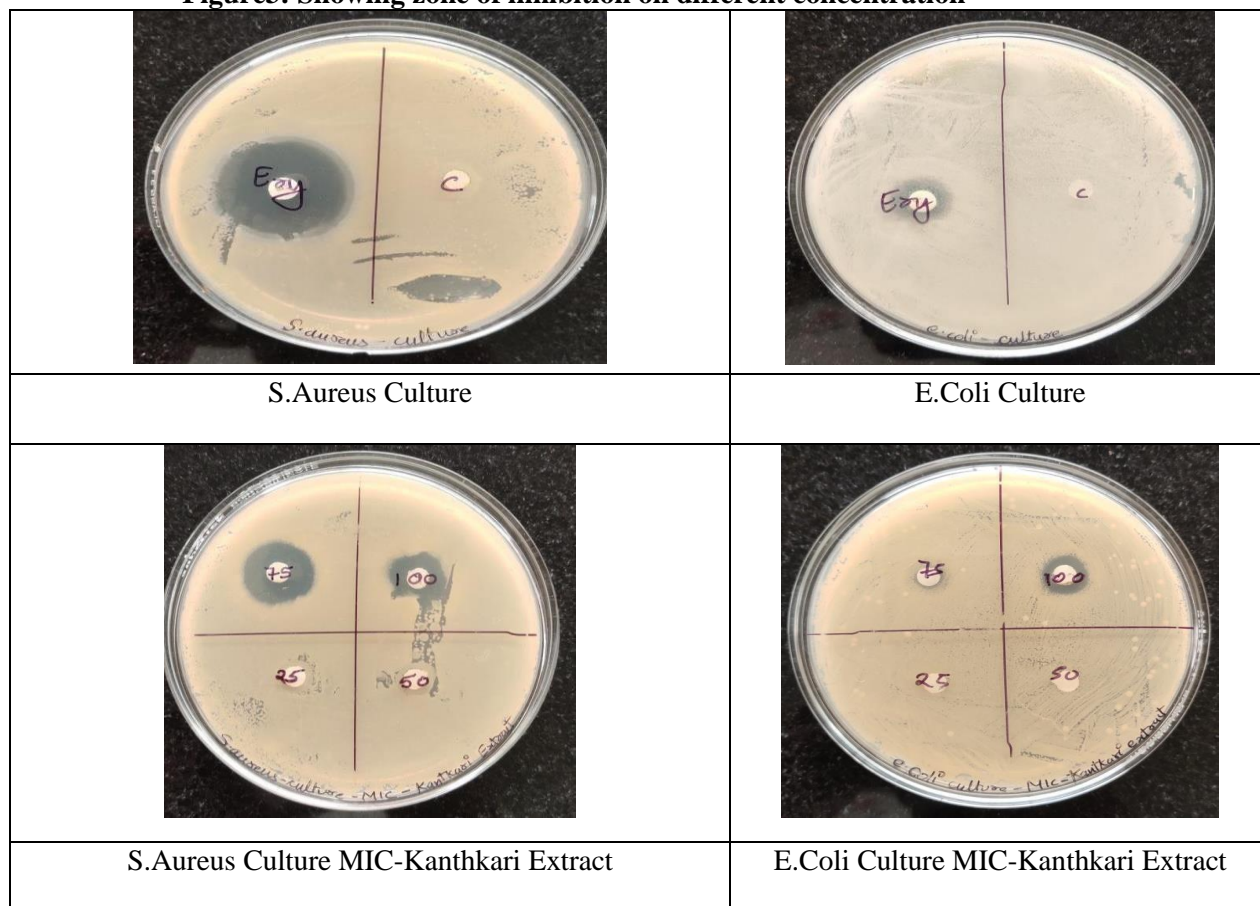
Standard Erythromycin – 15mcg.

Herbal Drug concentrations used are:

- a) Stock Solution – 500mg/1000µl
- b) 100mg/500µl
- c) 65mg/500µl
- d) 50mg/500µl
- e) 25mg/500µl

The relative effectiveness of a compound was determined by comparing the diameter of the zone of inhibition of bacterial growth around the discs. The Antibacterial activity of plant extracts and formulation with respect to Erythromycin antibiotic disc is mentioned in the table 7:

Figure 3: Showing zone of inhibition on different concentration



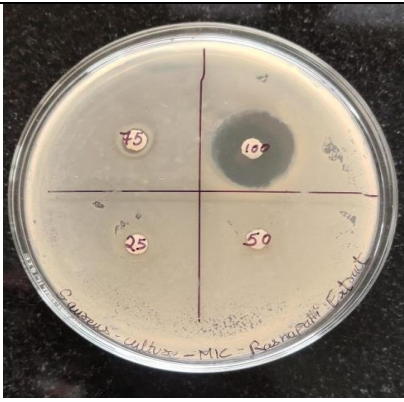
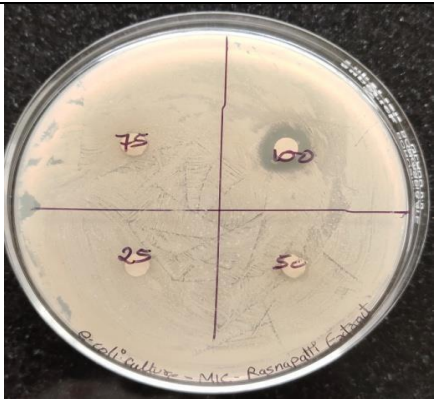
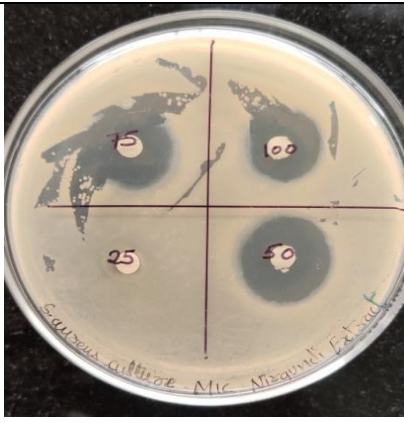
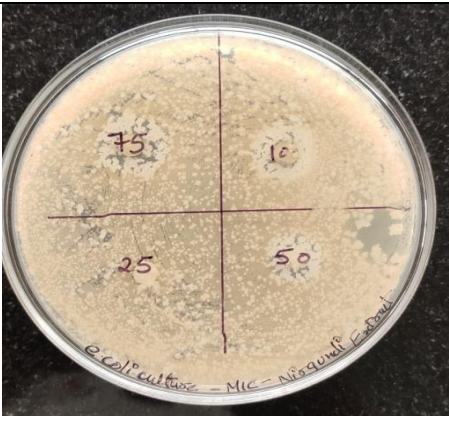

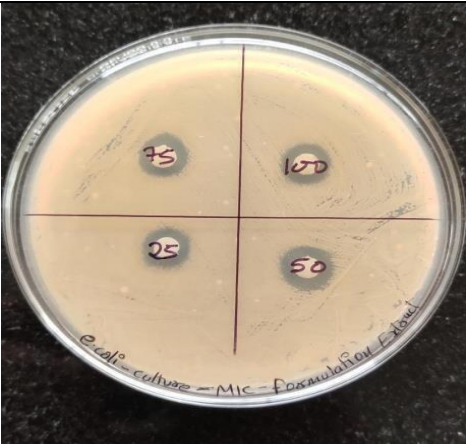
| | |
|---|--|
|  |  |
| S.Aureus Culture MIC-RasnapattiExtract | E.Coli Culture MIC-Rasnapatti Extract |
|  |  |
| S.Aureus Culture MIC-Nirgundi Extract | E.Coli Culture MIC-Nirgundi Extract |
|  |  |
| S.Aureus Culture MIC-Formulation | E.Coli Culture MIC-Formulation |

Table 7: Effect of extract of rasnapatti and Erythromycin on Zone of inhibition on different concentration

| Name of extract | Name of micro-organisms | Zone of inhibition on different concentration | | | | | |
|--------------------|------------------------------|---|---------|---------|---------|---------|----------|
| | | Erythromycin | Control | 25mg/ml | 50mg/ml | 65mg/ml | 100mg/ml |
| Rasnapatti extract | <i>Staphylococcus aureus</i> | 12 mm | 0 | 0 | 0 | 1 mm | 8 mm |
| | <i>E. coli</i> | 2 mm | 0 | 0 | 0 | 0 | 2 mm |

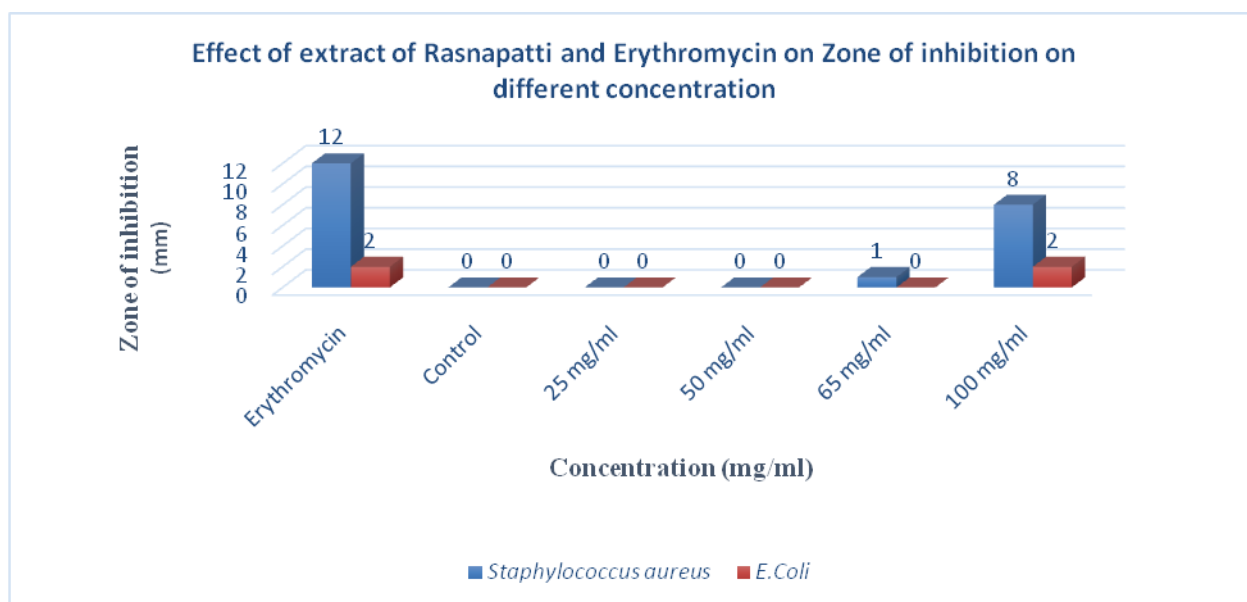


Figure 4: Effect of extract of rasnapatti and Erythromycin on Zone of inhibition on different concentration

Table 8: Effect of extract of Kantkari and Erythromycin on Zone of inhibition on different concentration

| Name of extract | Name of micro-organisms | Zone of inhibition on different concentration | | | | | |
|------------------|------------------------------|---|---------|---------|---------|---------|----------|
| | | Erythromycin | Control | 25mg/ml | 50mg/ml | 65mg/ml | 100mg/ml |
| Kantkari extract | <i>Staphylococcus aureus</i> | 12 mm | 0 | 0 | 0 | 6 mm | 2 mm |
| | <i>E. coli</i> | 2 mm | 0 | 0 | 0 | 1 mm | 2.2 mm |

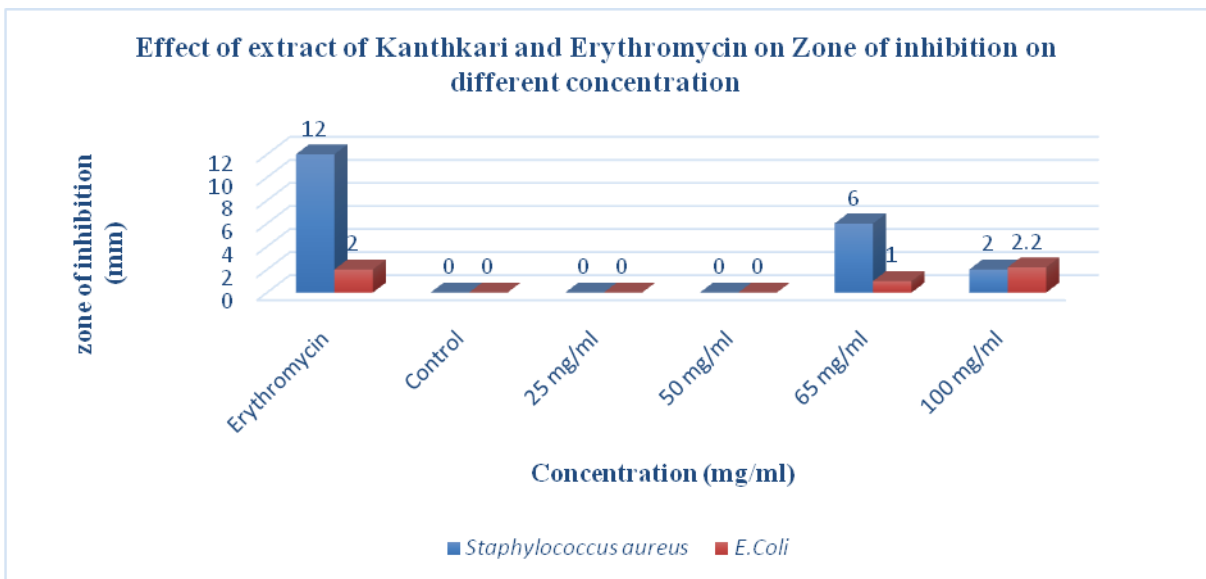


Figure 5: Effect of extract of Kantkari and Erythromycin on Zone of inhibition on different concentration

Table 9: Effect of extract of Nirgundi and Erythromycin on Zone of inhibition on different concentration

| Name of extract | Name of micro-organisms | Zone of inhibition on different concentration | | | | | |
|------------------|------------------------------|---|---------|---------|---------|---------|----------|
| | | Erythromycin | Control | 25mg/ml | 50mg/ml | 65mg/ml | 100mg/ml |
| Nirgundi extract | <i>Staphylococcus aureus</i> | 12 mm | 0 | 0 | 6.5 mm | 9 mm | 8 mm |
| | <i>E. coli</i> | 2 mm | 0 | 0 | 2 mm | 5.5 mm | 5 mm |

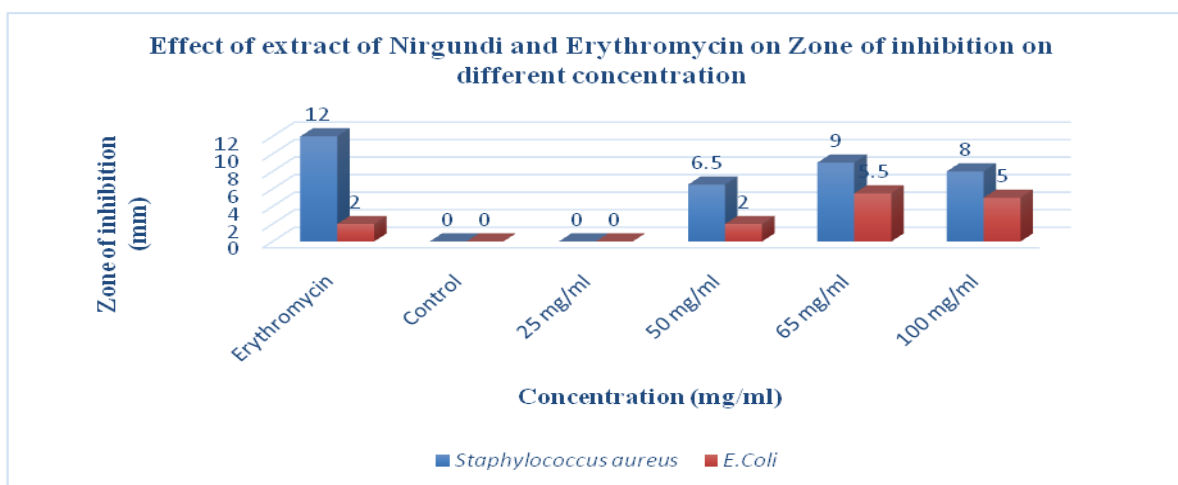


Fig6: Effect of extract of Nirgundi and Erythromycin on Zone of inhibition on different concentration

Table 10: Effect of extract of Formulation and Erythromycin on Zone of inhibition on different concentration

| Name of extract | Name of micro-organisms | Zone of inhibition on different concentration | | | | | |
|-----------------|------------------------------|---|---------|---------|---------|---------|----------|
| | | Erythromycin | Control | 25mg/ml | 50mg/ml | 65mg/ml | 100mg/ml |
| Formulation | <i>Staphylococcus aureus</i> | 12 mm | 0 | 5 mm | 5.5 mm | 6 mm | 6 mm |
| | <i>E. coli</i> | 2 mm | 0 | 1 mm | 2 mm | 2 mm | 2.5 mm |

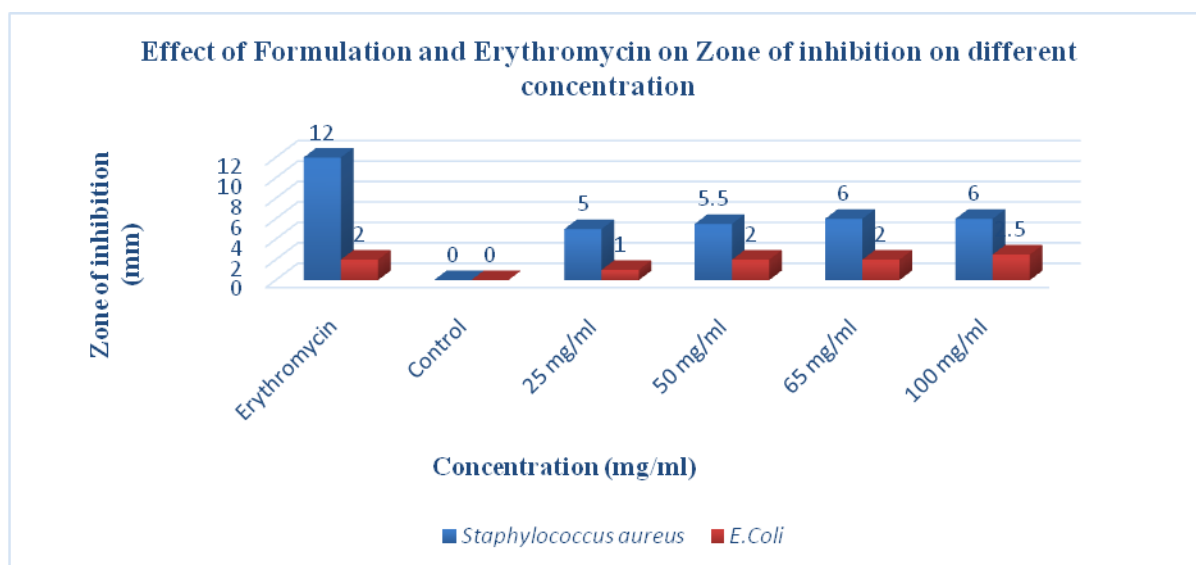


Fig 7: Effect of extract of Formulation and Erythromycin on Zone of inhibition on different concentration

Result and Discussion

The present study features a novel herbal skin care formulation with potential antibacterial properties. Prepared herbal cream contains combinations of herbal ingredients that are formulated to combat bacterial infection and reduce oxidative stress. This work deals with description, review of literature, pharmacognostical evaluation of plant material, preliminary phytochemical studies, thin layer chromatography, preparation of herbal cream comprising three plant extracts and in vitro antibacterial activity of extracts and formulation and pharmaceutical evaluation of formulations.

Analytical study of plant material

In recent years there has been an emphasis on standardization of medicinal plants of therapeutic potential. Identification and evaluation of plant drugs by pharmacognostic studies are very reliable, accurate and inexpensive means. According to World Health Organization (WHO) these studies are the first step towards establishing its identity and purity. This study was focused on the physical and chemical characters of roughly ground plant of Rasna, Kanthkari and Nirgundi. Study started with the analysis of physical parameters of above-mentioned plants; in this, we studied Loss on drying, Total ash value, Acid insoluble ash value, Water soluble ash value,

Alcohol extractive value and Water extractive value. Preliminary physical characters of different extracts, qualitative chemical tests and TLC was performed to get an idea of presence of different classes of secondary metabolites, which are useful indicators of both efficacy and potential toxicity. The results of total ash value, acid insoluble ash value, water soluble ash values help in detected the extent of adulteration as well as establish the quality and purity of the drug. The solvent used for the extraction dissolves appreciable quantities of desired constituents. The selectivity of the solvent is important not only for the yield of one or more principal substances, but also for the qualitative and quantitative composition of the accompanying substances. Moreover, extractive values are primarily useful for the determination of exhausted or adulterated drugs. From the present study, it can be concluded that aqueous extractive values of selected medicinal plants were higher than the ethanol solvents. The result suggests that large quantity of polar soluble phytoconstituents were present in crude extracts.

TLC profiling of ethanol extracts gives an impressive result that directing towards the presence standard Quercetin. The TLC method is the best choice for the identification of the secondary metabolite present in plants. The qualitative as well as quantitative HPTLC analysis under standard conditions provides chromatograms, which are very useful for controlling the quality of the phyto-pharmaceuticals. These studies provided referential information in regard to its identification parameters assumed significantly in the way of acceptability of herbal drugs in present scenario of lack of regulatory laws to control quality of herbal drugs.

In vitro antibacterial activity

Antibacterial activity was done using ethanolic extract by disc diffusion method at different concentration against two bacterial strains showed varied level of zone of inhibition ranged from 1mm-9mm. Formulated cream shows significant result against *S.aureus* and *E.coli* thereby establishing promising dosage form .

Due to Restrictions of CPCSEA for the use of animals in experimental pharmacological research, such as ethical issues and the lack of rationale for their use when other suitable

methods are available. Hence, in the present study the assessment of the antimicrobial property of the herbal extract of Kanthkari, Rasana, and Nirgundi and formulation done by disc diffusion method at different concentration against two bacterial strains showed varied level of zone of inhibition.

Conclusion

The plant products over the synthetic compound in the treatment of diseases are needed because it does not have a deleterious effect in higher plants and animals including man. The urge to research new drugs from natural sources is now moving out of the herbalist's shop, away from the core texts into the drug research laboratories. India is home to a variety of traditional medicine systems that rely largely on native plant species for their raw drug materials. Therefore, now there is a need to look back toward traditional medicines, which can serve as a novel therapeutic agent. After getting information from various Vaidhyas, Ayurvedic doctors, and from a literature survey, it was found that *Kanthkari*, *Rasana*, and *Nirgundi* was used since ancient times to treat various diseases and prone to have antimicrobial properties. Hence, O/W emulsion-based cream formulation composed of *Kanthkari*, *Rasana*, and *Nirgundi* prepared further HPTLC profiling and Antibacterial properties was evaluated. HPTLC profiling confirms the presence of Quercetin in formulation and extract with an Rf of 0.22. Antibacterial activity established by disk diffusion method shows 6mm and 2.5 mm the zone of inhibition by O/W emulsion-based cream formulation against *S.aureus* and *E.coli* respectively. It is concluded that the cream formulation exhibits significant antibacterial activity. The results authenticate the fact that prepared herbal skin care formulations could be used safely and effectively and are good alternatives for bacterial infection.

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